

Nucleotide Metabolism 22



For additional ancillary materials related to this chapter, please visit [thePoint](#).

I. OVERVIEW

Ribonucleoside and deoxyribonucleoside phosphates (nucleotides) are essential for all cells. Without them, neither ribonucleic acid (RNA) nor deoxyribonucleic acid (DNA) can be produced, and, therefore, proteins cannot be synthesized or cells proliferate. Nucleotides also serve as carriers of activated intermediates in the synthesis of some carbohydrates, lipids, and conjugated proteins (for example, uridine diphosphate [UDP]-glucose and cytidine diphosphate [CDP]-choline) and are structural components of several essential coenzymes, such as coenzyme A, flavin adenine dinucleotide (FAD[H₂]), nicotinamide adenine dinucleotide (NAD[H]), and nicotinamide adenine dinucleotide phosphate (NADP[H]). Nucleotides, such as cyclic adenosine monophosphate (cAMP) and cyclic guanosine monophosphate (cGMP), serve as second messengers in signal transduction pathways. In addition, nucleotides play an important role as energy sources in the cell. Finally, nucleotides are important regulatory compounds for many of the pathways of intermediary metabolism, inhibiting or activating key enzymes. The purine and pyrimidine bases found in nucleotides can be synthesized de novo or can be obtained through salvage pathways that allow the reuse of the preformed bases resulting from normal cell turnover. [Note: Little of the purines and pyrimidines supplied by diet is utilized and is degraded instead.]

II. STRUCTURE

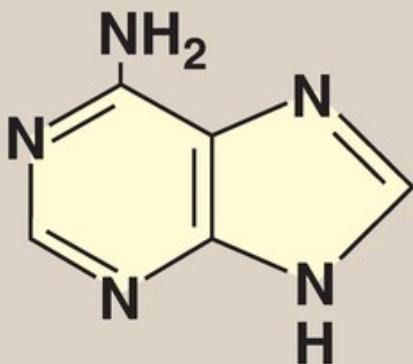
Nucleotides are composed of a nitrogenous base; a pentose monosaccharide; and one, two, or three phosphate groups. The nitrogen-containing bases belong to

two families of compounds: the purines and the pyrimidines.

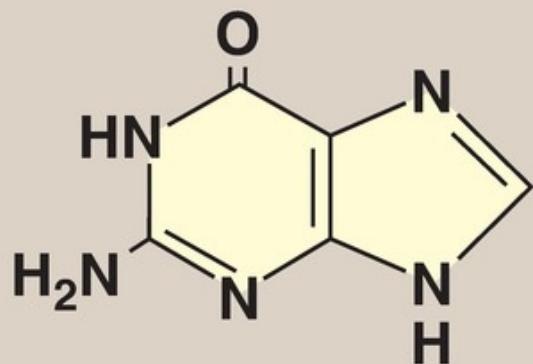
A. Purine and pyrimidine bases

Both DNA and RNA contain the same purine bases: adenine (A) and guanine (G). Both DNA and RNA contain the pyrimidine cytosine (C), but they differ in their second pyrimidine base: DNA contains thymine (T), whereas RNA contains uracil (U). T and U differ in that only T has a methyl group ([Fig. 22.1](#)). Unusual (modified) bases are occasionally found in some species of DNA (for example, in some viral DNA) and RNA (for example, in transfer RNA [tRNA]). Base modifications include methylation, glycosylation, acetylation, and reduction. Some examples of unusual bases are shown in [Figure 22.2](#). [Note: The presence of an unusual base in a nucleotide sequence may aid in its recognition by specific enzymes or protect it from being degraded by **nucleases**.]

DNA and RNA Purines

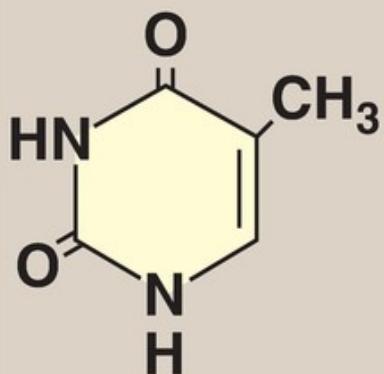


Adenine (A)

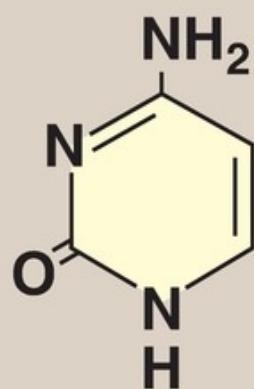


Guanine (G)

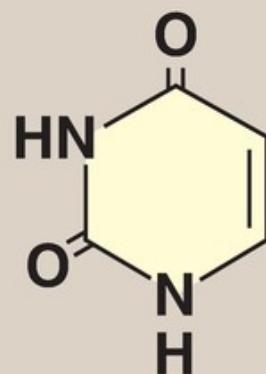
RNA Pyrimidines



Thymine (T)



Cytosine (C)

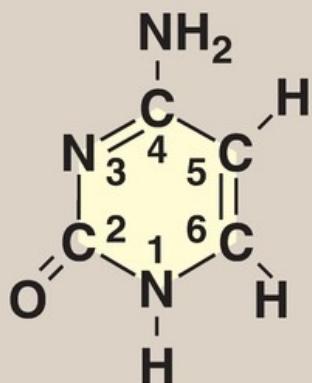


Uracil (U)

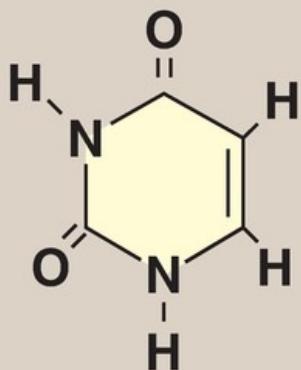
DNA Pyrimidines

Figure 22.1 Purines and pyrimidines commonly found in DNA and RNA.

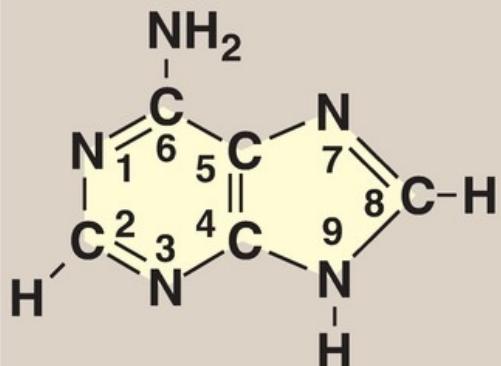
Common Base



Cytosine

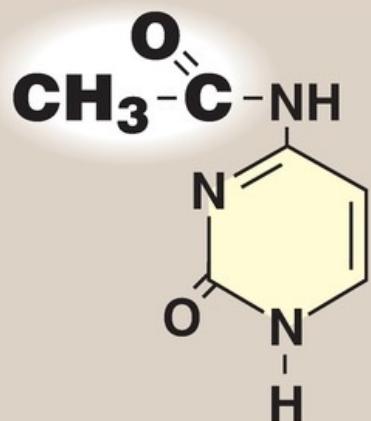


Uracil

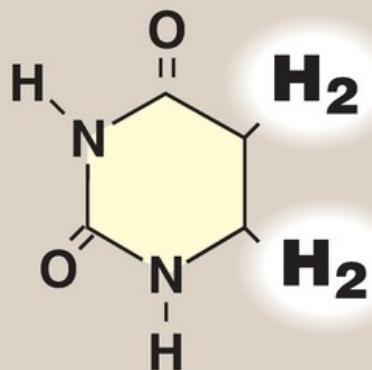


Adenine

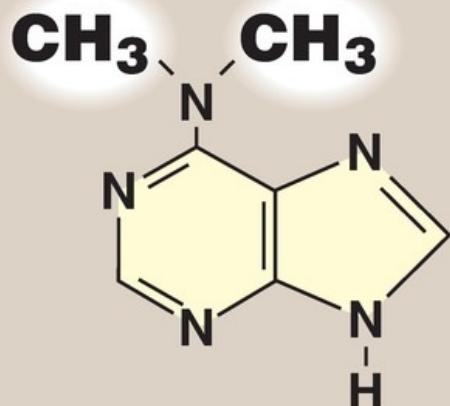
Unusual Base



N⁴-Acetylcytosine



Dihydrouracil



N⁶,N⁶-Dimethyladenine

Figure 22.2 Examples of unusual bases.

B. Nucleosides

The addition of a pentose sugar to a base through an N-glycosidic bond (see p. 86) produces a nucleoside. If the sugar is ribose, a ribonucleoside is produced, and if the sugar is 2-deoxyribose, a deoxyribonucleoside is produced ([Fig. 22.3A](#)). The ribonucleosides of A, G, C, and U are named adenosine, guanosine, cytidine, and uridine, respectively. The deoxyribonucleosides of A, G, C, and T have the added prefix deoxy- (for example, deoxyadenosine). [Note: The compound deoxythymidine is often simply called thymidine, with the deoxy- prefix being understood, because it is incorporated into DNA only.] The carbon and nitrogen atoms in the rings of the base and the sugar are numbered separately (see [Fig. 22.3B](#)). [Note: Carbons in the pentose are numbered 1' to 5'. Thus, when the 5'-carbon of a nucleoside (or nucleotide) is referred to, a carbon atom in the pentose, rather than an atom in the base, is being specified.]

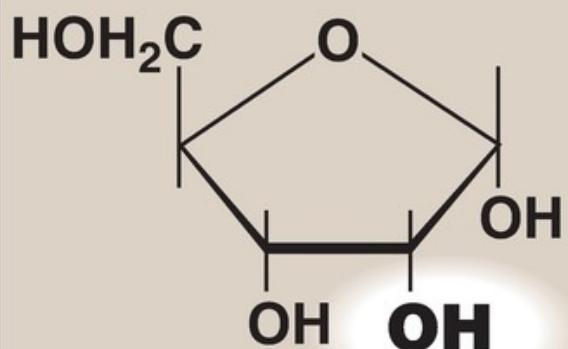
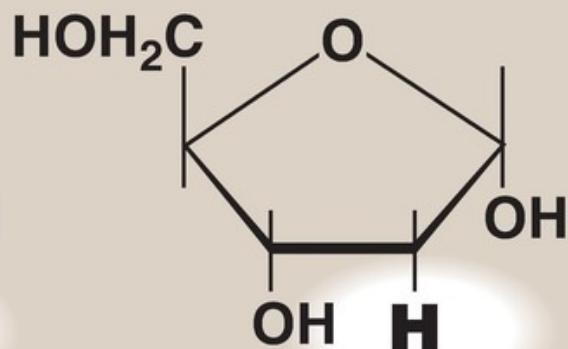
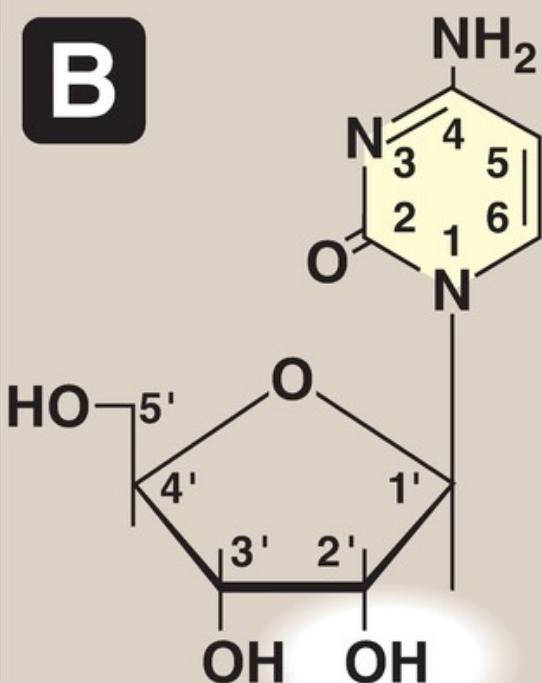
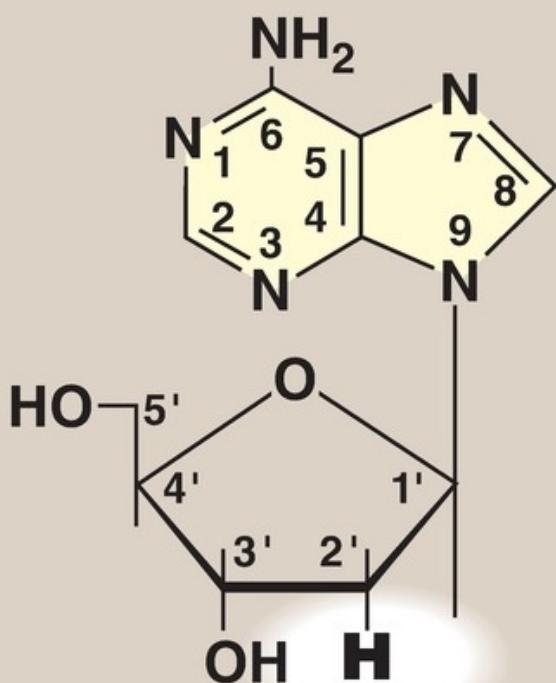
A**RNA****DNA****Ribose****2-Deoxyribose****B****Cytidine****Deoxyadenosine**

Figure 22.3 A. Pentoses found in nucleic acids. B. Examples of the numbering systems for purine- and pyrimidine-containing nucleosides.

C. Nucleotides

The addition of one or more phosphate groups to a nucleoside produces a nucleotide. The first phosphate group is attached by an ester linkage to the 5'-OH of the pentose, forming a nucleoside 5'-phosphate or a 5'-nucleotide. The type of pentose is denoted by the prefix in the names 5'-ribonucleotide and 5'-deoxyribonucleotide. If one phosphate group is attached to the 5'-carbon of the pentose, the structure is a nucleoside monophosphate, like adenosine monophosphate (AMP, or adenylate). If a second or third phosphate is added to the nucleoside, a nucleoside diphosphate (for example, adenosine diphosphate [ADP] or triphosphate, for example, ATP) results ([Fig. 22.4](#)). The second and third phosphates are each connected to the nucleotide by a “high-energy bond” (a bond with a large, negative change in free energy [$-\Delta G$, see p. 70] of hydrolysis). [Note: The phosphate groups are responsible for the negative charges associated with nucleotides and cause DNA and RNA to be referred to as nucleic acids.]

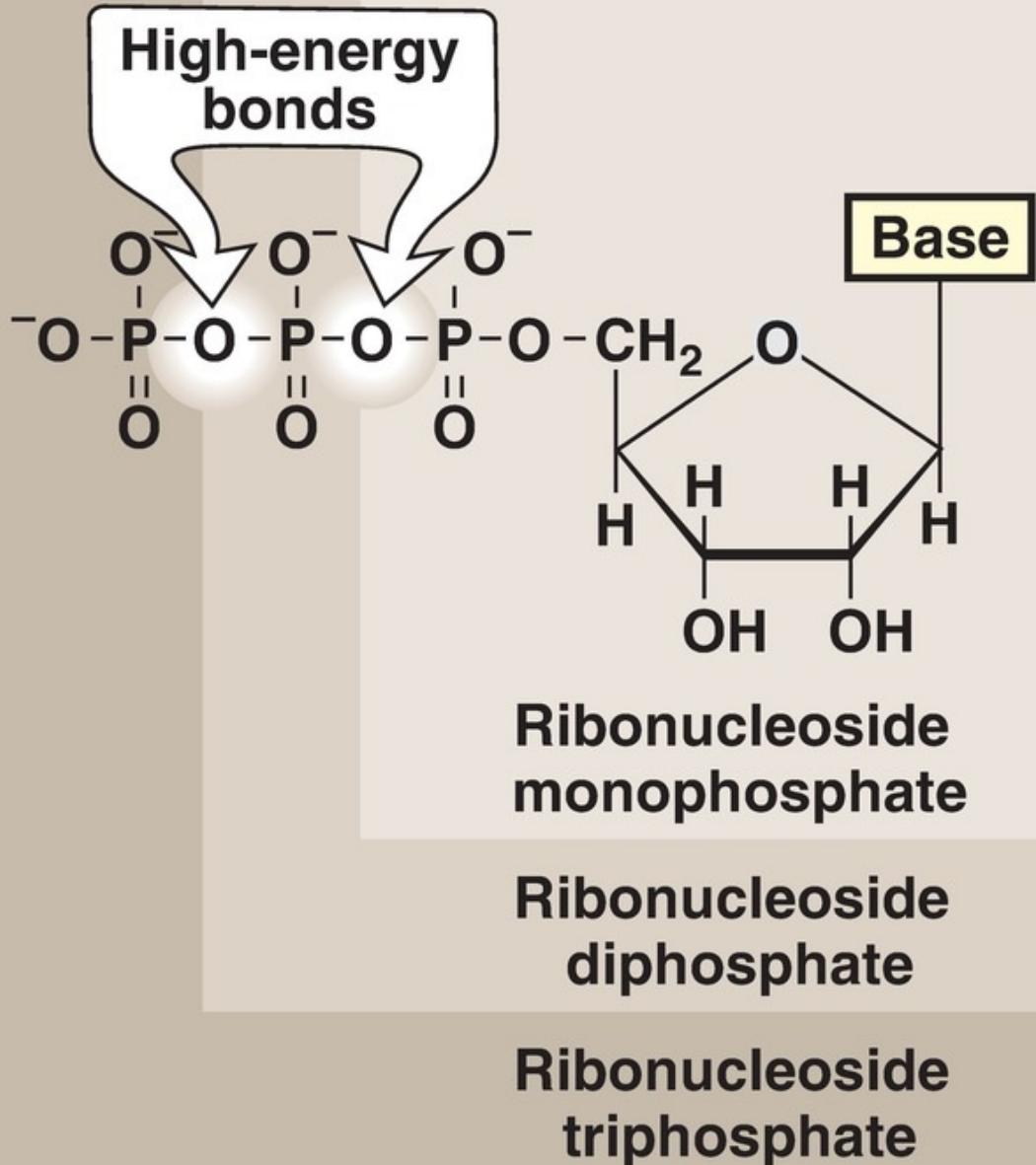


Figure 22.4 Ribonucleoside monophosphate, diphosphate, and triphosphate.

III. PURINE NUCLEOTIDE SYNTHESIS

The atoms of the purine ring are contributed by a number of compounds, including amino acids (aspartate, glycine, and glutamine), carbon dioxide (CO_2), and N^{10} -formyltetrahydrofolate (N^{10} -formyl-THF), as shown in [Figure 22.5](#). The purine ring is constructed primarily in the liver by a series of reactions that add

the donated carbons and nitrogens to a preformed ribose 5-phosphate. [Note: Synthesis of ribose 5-phosphate from glucose 6-phosphate by the pentose phosphate pathway is discussed on p. 147.]

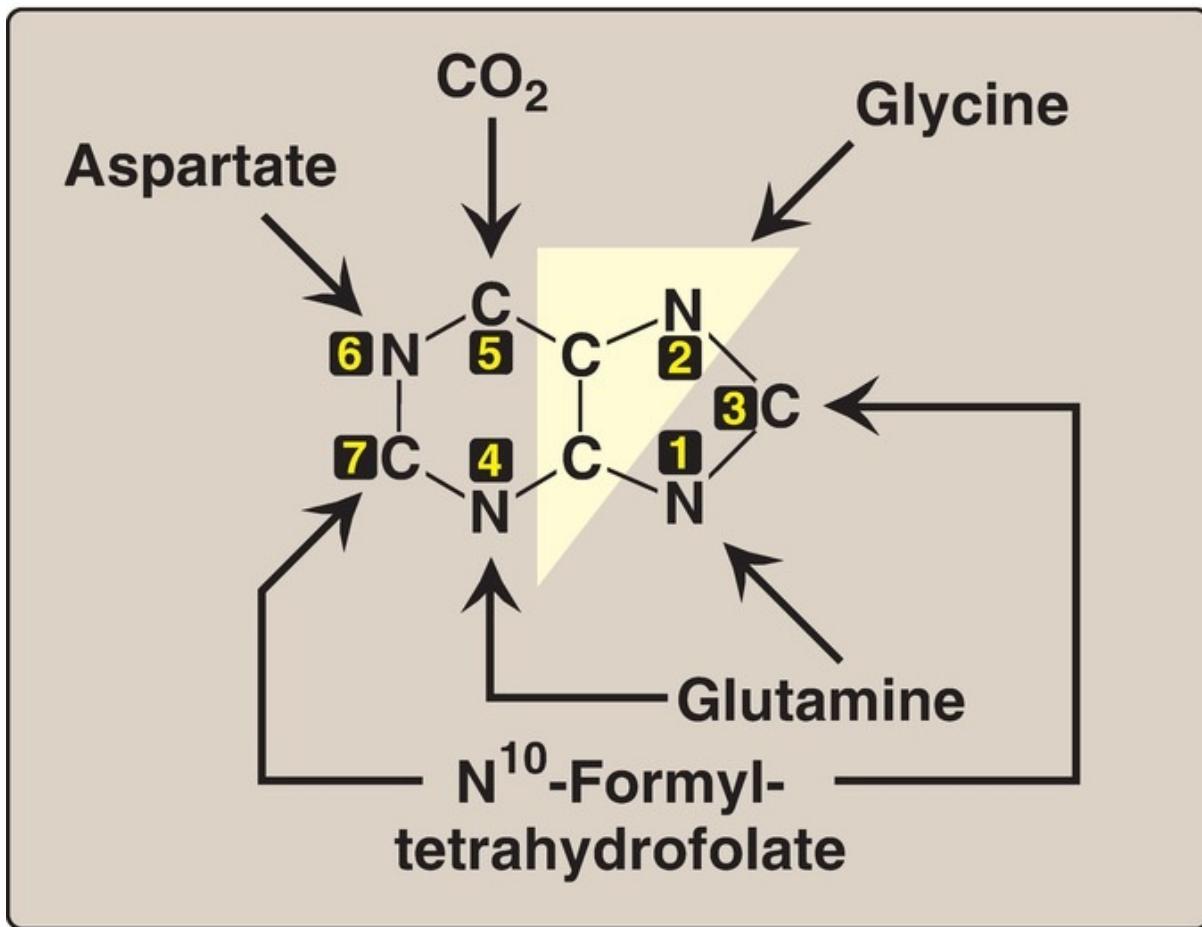


Figure 22.5 Sources of the individual atoms in the purine ring. The order in which the atoms are added is shown by the numbers in the black boxes (see Fig. 22.7). CO₂ = carbon dioxide.

A. 5-Phosphoribosyl-1-pyrophosphate synthesis

5-Phosphoribosyl-1-pyrophosphate (PRPP) is an activated pentose that participates in the synthesis and salvage of purines and pyrimidines. Synthesis of PRPP from ATP and ribose 5-phosphate is catalyzed by **PRPP synthetase** (Fig. 22.6). This X-linked enzyme is activated by inorganic phosphate and inhibited by purine nucleotides (end-product inhibition). [Note: Because the sugar moiety of PRPP is ribose, ribonucleotides are the

end products of *de novo* purine synthesis. When deoxyribonucleotides are required for DNA synthesis, the ribose sugar moiety is reduced (see p. 297).]

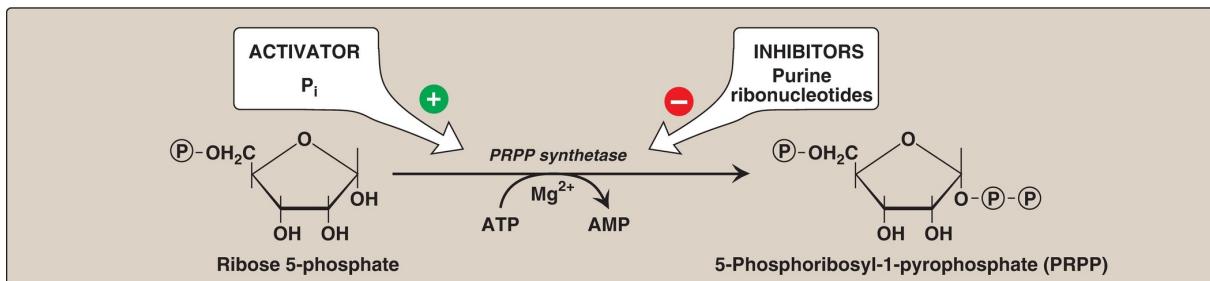


Figure 22.6 Synthesis of PRPP, showing the activator and inhibitors of the reaction. [Note: This is not the committed step of purine synthesis because PRPP is used in other pathways such as salvage (see p. 296).] = phosphate; P_i = inorganic phosphate; AMP = adenosine monophosphate; Mg = magnesium.

B. 5-Phosphoribosylamine synthesis

Synthesis of 5-phosphoribosylamine from PRPP and glutamine is shown in [Figure 22.7](#). The amide group of glutamine replaces the pyrophosphate group attached to carbon 1 of PRPP. This is the committed step in purine nucleotide biosynthesis. The enzyme that catalyzes the reaction, **glutamine:phosphoribosylpyrophosphate amidotransferase (GPAT)**, is inhibited by the purine 5'-nucleotides AMP and guanosine monophosphate (GMP, or guanylate), the end products of the pathway. The rate of the reaction is also controlled by the intracellular concentration of PRPP. [Note: The concentration of PRPP is normally far below the Michaelis constant (K_m) for the **GPAT**. Therefore, any small change in the PRPP concentration causes a proportional change in rate of the reaction (see p. 59).]

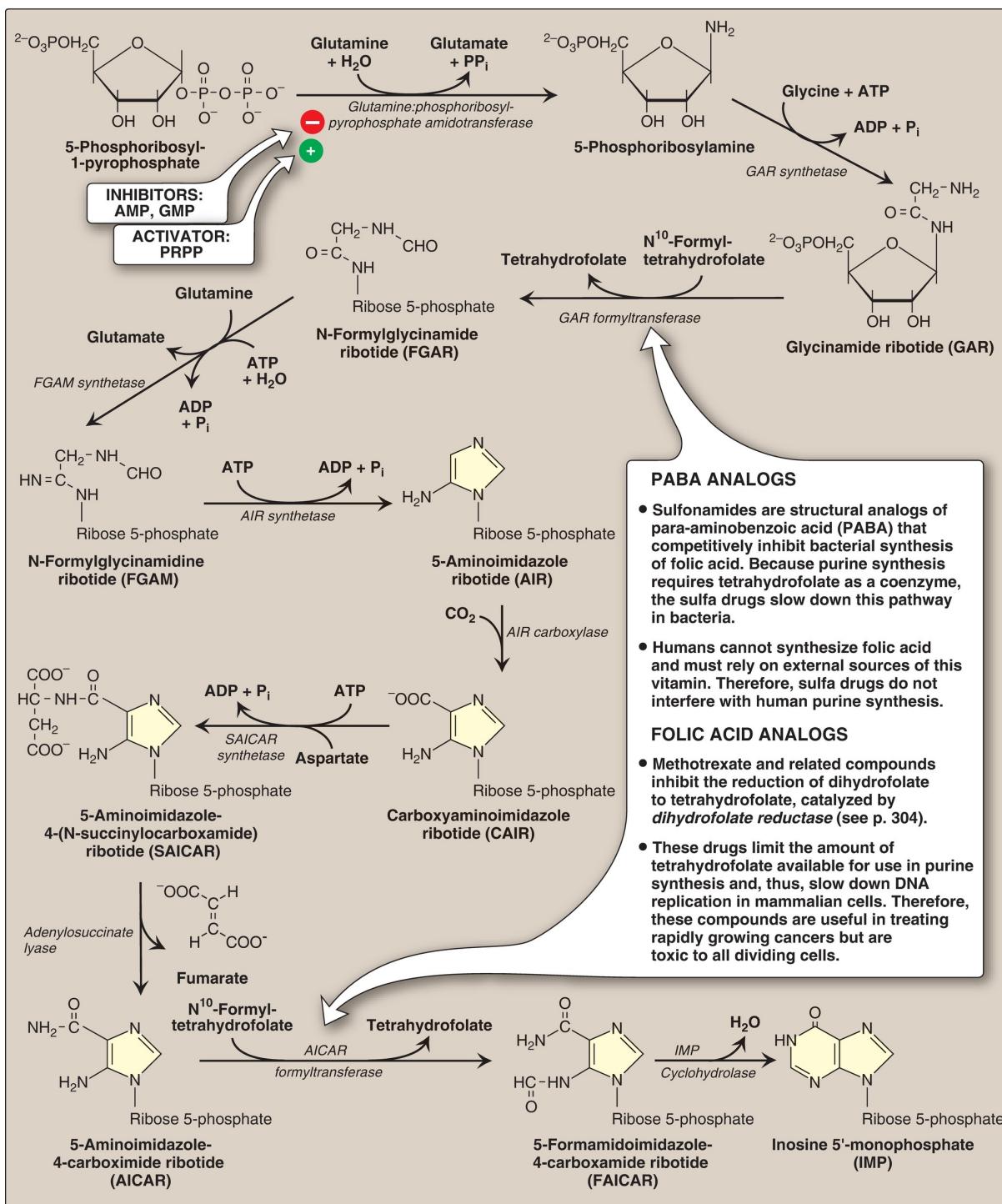


Figure 22.7 *De novo* synthesis of purine nucleotides, showing the inhibitory effect of some structural analogs. AMP and ADP = adenosine mono- and diphosphates; GMP = guanosine monophosphate; PRPP = 5-phosphoribosyl-1-pyrophosphate; P_i = inorganic phosphate; PP_i = pyrophosphate; CO₂ = carbon dioxide.

C. Inosine monophosphate synthesis

The next nine steps in purine nucleotide biosynthesis leading to the synthesis of inosine monophosphate ([IMP] whose base is hypoxanthine) are illustrated in [Figure 22.7](#). IMP is the parent purine nucleotide for AMP and GMP. Four steps in this pathway require ATP as an energy source, and two steps in the pathway require N¹⁰-formyl-THF as a one-carbon donor (see p. 267). [Note: Hypoxanthine is found in tRNA (see Fig. 32.9 on p. 453).]

D. Synthetic inhibitors

Some synthetic inhibitors of purine synthesis (for example, the sulfonamides) are designed to inhibit the growth of rapidly dividing microorganisms without interfering with human cell functions (see [Fig. 22.7](#)). Other purine synthesis inhibitors, such as structural analogs of folic acid (for example, methotrexate), are used pharmacologically to control the spread of cancer by interfering with the synthesis of nucleotides and, therefore, of DNA and RNA (see [Fig. 22.7](#)).

Inhibitors of human purine synthesis are extremely toxic to tissues, especially to developing structures such as in a fetus, or to cell types that normally replicate rapidly, including those of bone marrow, skin, gastrointestinal (GI) tract, immune system, or hair follicles. As a result, individuals taking such anticancer drugs can experience adverse effects, including anemia, scaly skin, GI tract disturbance, immunodeficiency, and hair loss.

E. Adenosine and guanosine monophosphate synthesis

The conversion of IMP to either AMP or GMP uses a two-step, energy- and nitrogen-requiring pathway ([Fig. 22.8](#)). [Note: AMP synthesis requires guanosine triphosphate (GTP) as an energy source and aspartate as a nitrogen source, whereas GMP synthesis requires ATP and glutamine.] Also, the first reaction in each pathway is inhibited by the end product of that pathway. This provides a mechanism for diverting IMP to the synthesis

of the purine present in lesser amounts. If both AMP and GMP are present in adequate amounts, the de novo pathway of purine nucleotide synthesis is inhibited at the **GPAT** step.

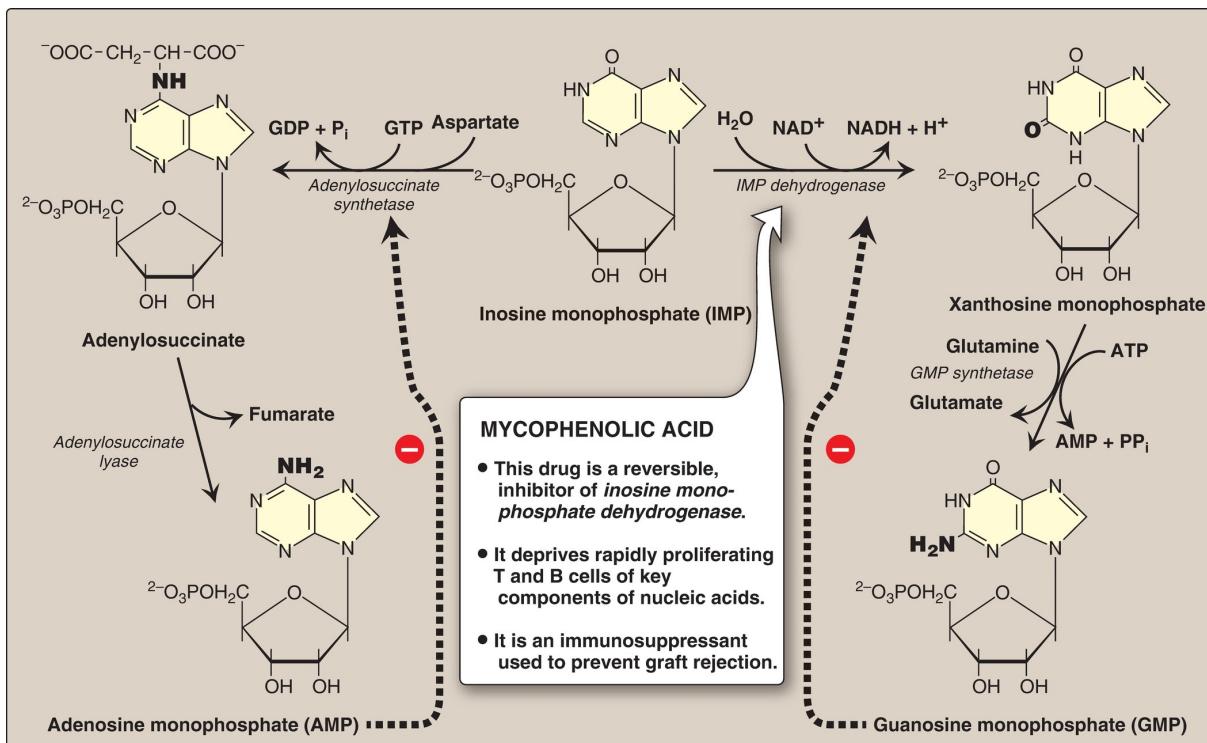


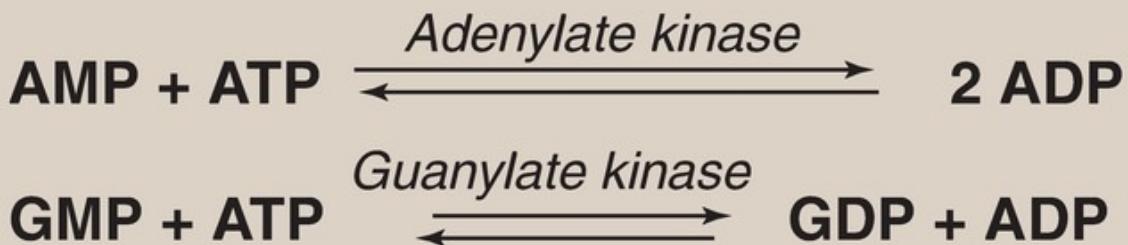
Figure 22.8 Conversion of IMP to AMP (or, adenylate) and GMP (or, guanylate) showing feedback inhibition. NAD(H) = nicotinamide adenine dinucleotide; GDP and GTP = guanosine di- and triphosphates; P_i = inorganic phosphate; PP_i = pyrophosphate.

F. Nucleoside di- and triphosphate synthesis

Nucleoside diphosphates are synthesized from the corresponding nucleoside monophosphates by base-specific **nucleoside monophosphate kinases** (Fig. 22.9). [Note: These **kinases** do not discriminate between ribose or deoxyribose in the substrate.] ATP is generally the source of the transferred phosphate because it is present in higher concentrations than the other nucleoside triphosphates. **Adenylate kinase** is particularly active in the liver and in muscle, where the turnover of energy from ATP is high. Its function is to maintain equilibrium among the adenine nucleotides (AMP, ADP, and ATP). Nucleoside diphosphates and triphosphates are interconverted by

nucleoside diphosphate kinase, an enzyme that, unlike the **monophosphate kinases**, has broad substrate specificity.

Base-specific nucleoside monophosphate kinases



Nucleoside diphosphate kinase

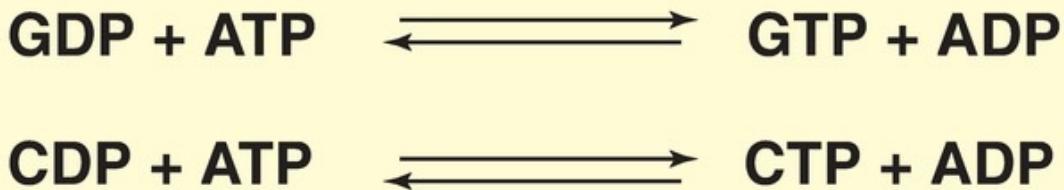


Figure 22.9 Conversion of nucleoside monophosphates to di- and triphosphates. AMP and ADP = adenosine mono- and diphosphates; GMP, GDP, and GTP = guanosine mono-, di-, and triphosphates; CDP and CTP = cytidine di- and triphosphates.

G. Purine salvage pathway

Purines that result from the normal turnover of cellular nucleic acids, or the small amount that is obtained from the diet and not degraded, can be converted to nucleoside triphosphates and used by the body. This is referred to as the salvage pathway for purines. [Note: Salvage is particularly important in the brain.]

1. purine base salvage to nucleotides: Two enzymes are involved: **adenine phosphoribosyltransferase (APRT)** and X-linked **hypoxanthine-**

guanine phosphoribosyltransferase (HGPRT). Both use PRPP as the source of the ribose 5-phosphate group (Fig. 22.10). The release of pyrophosphate and its subsequent hydrolysis by **pyrophosphatase** makes these reactions irreversible. [Note: Adenosine is the only purine nucleoside to be salvaged. It is phosphorylated to AMP by **adenosine kinase**.]

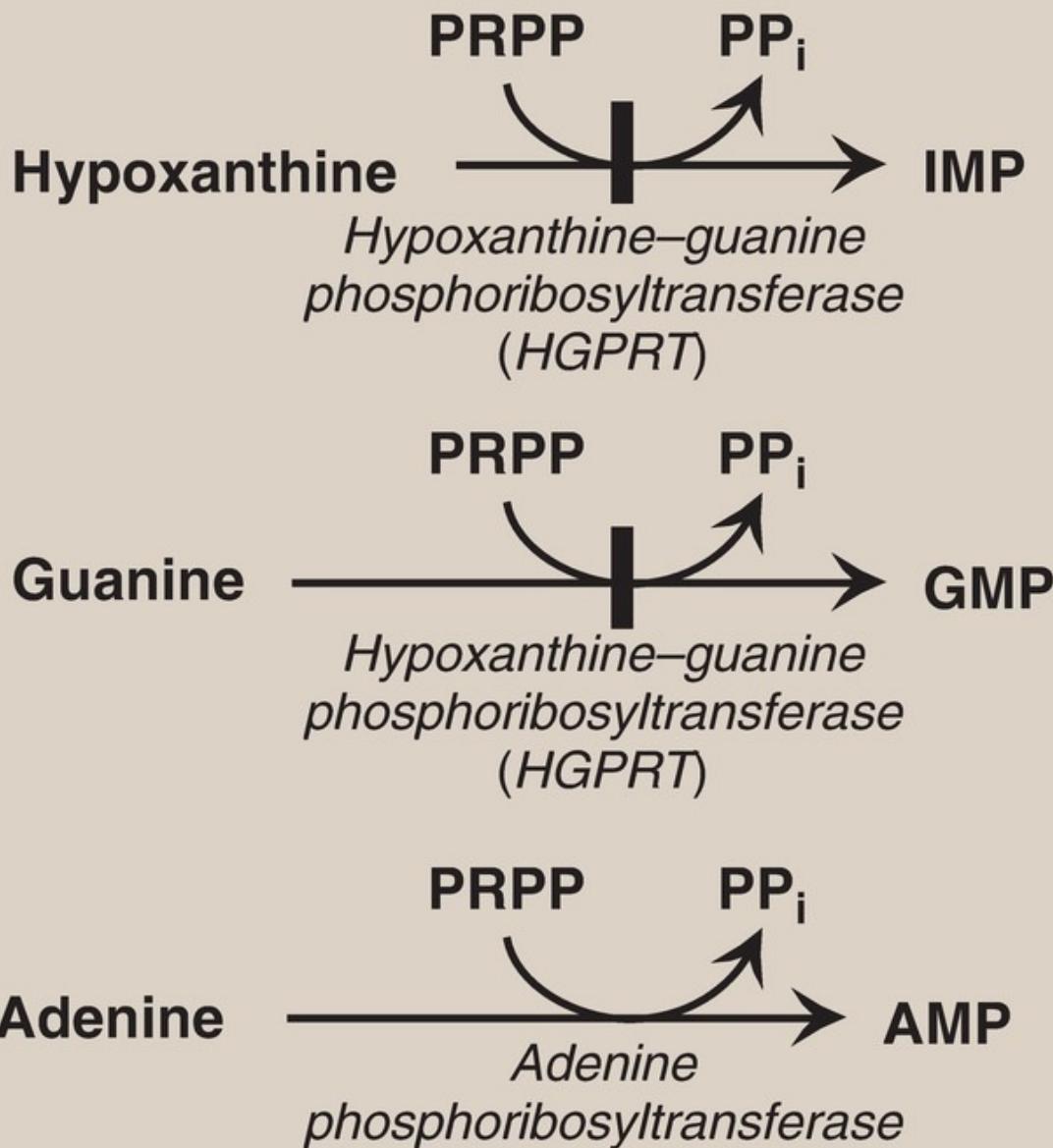


Figure 22.10 Salvage pathways of purine nucleotide synthesis. [Note: Virtually complete deficiency of **HGPRT** results in Lesch-Nyhan syndrome. Partial

deficiencies of **HGPRT** are known. As the amount of functional enzyme increases, the severity of the symptoms decreases.] IMP, GMP, and AMP = inosine, guanosine, and adenosine monophosphates; PRPP = 5-phosphoribosyl-1-pyrophosphate; PP_i = pyrophosphate.

2. Lesch-Nyhan syndrome: This is a rare, X-linked recessive disorder associated with a virtually complete deficiency of **HGPRT**. The deficiency results in an inability to salvage hypoxanthine or guanine, from which excessive amounts of uric acid, the end product of purine degradation, are then produced (see p. 298). In addition, the lack of this salvage pathway causes increased PRPP levels and decreased IMP and GMP levels. As a result, **GPAT** (the regulated step in purine synthesis) has excess substrate and decreased inhibitors available, and de novo purine synthesis is increased. The combination of decreased purine reutilization and increased purine synthesis results in increased degradation of purines and the production of large amounts of uric acid, making **HGPRT** deficiency an inherited cause of hyperuricemia. In patients with Lesch-Nyhan syndrome, the hyperuricemia frequently results in the formation of uric acid stones in the kidneys (urolithiasis) and the deposition of urate crystals in the joints (gouty arthritis) and soft tissues. In addition, the syndrome is characterized by motor dysfunction, cognitive deficits, and behavioral disturbances that include self-mutilation (for example, biting of lips and fingers), as shown in [Figure 22.11](#).



Figure 22.11 Lesions on the lips of a patient with Lesch-Nyhan syndrome.

IV. DEOXYRIBONUCLEOTIDE SYNTHESIS

The nucleotides described thus far all contain ribose (ribonucleotides). DNA synthesis, however, requires 2'-deoxyribonucleotides, which are produced from ribonucleoside diphosphates by the enzyme **ribonucleotide reductase** during the S-phase of the cell cycle (see p. 423). [Note: The same enzyme acts on pyrimidine ribonucleotides.]

A. Ribonucleotide reductase

Ribonucleotide reductase (ribonucleoside diphosphate reductase) is a dimer composed of two nonidentical subunits, R1 (or, α) and the smaller R2 (or, β), and is specific for the reduction of purine nucleoside diphosphates (ADP and GDP) and pyrimidine nucleoside diphosphates (CDP and UDP) to their deoxy forms (dADP, dGDP, dCDP, and dUDP). The immediate donors of the hydrogen atoms needed for the reduction of the 2'-hydroxyl group are two sulfhydryl ($-SH$) groups on the enzyme itself (R1 subunit), which form a disulfide bond during the reaction (see p. 19). [Note: R2 contains the stable tyrosyl radical required for catalysis at R1.]

1. Reduced enzyme regeneration: In order for **ribonucleotide reductase** to continue to produce deoxyribonucleotides at R1, the disulfide bond created during the production of the 2'-deoxy carbon must be reduced. The source of the reducing equivalents is thioredoxin, a protein coenzyme of **ribonucleotide reductase**. Thioredoxin contains two cysteine residues separated by two amino acids in the peptide chain. The two $-SH$ groups of thioredoxin donate their hydrogen atoms to **ribonucleotide reductase**, forming a disulfide bond in the process ([Fig. 22.12](#)).

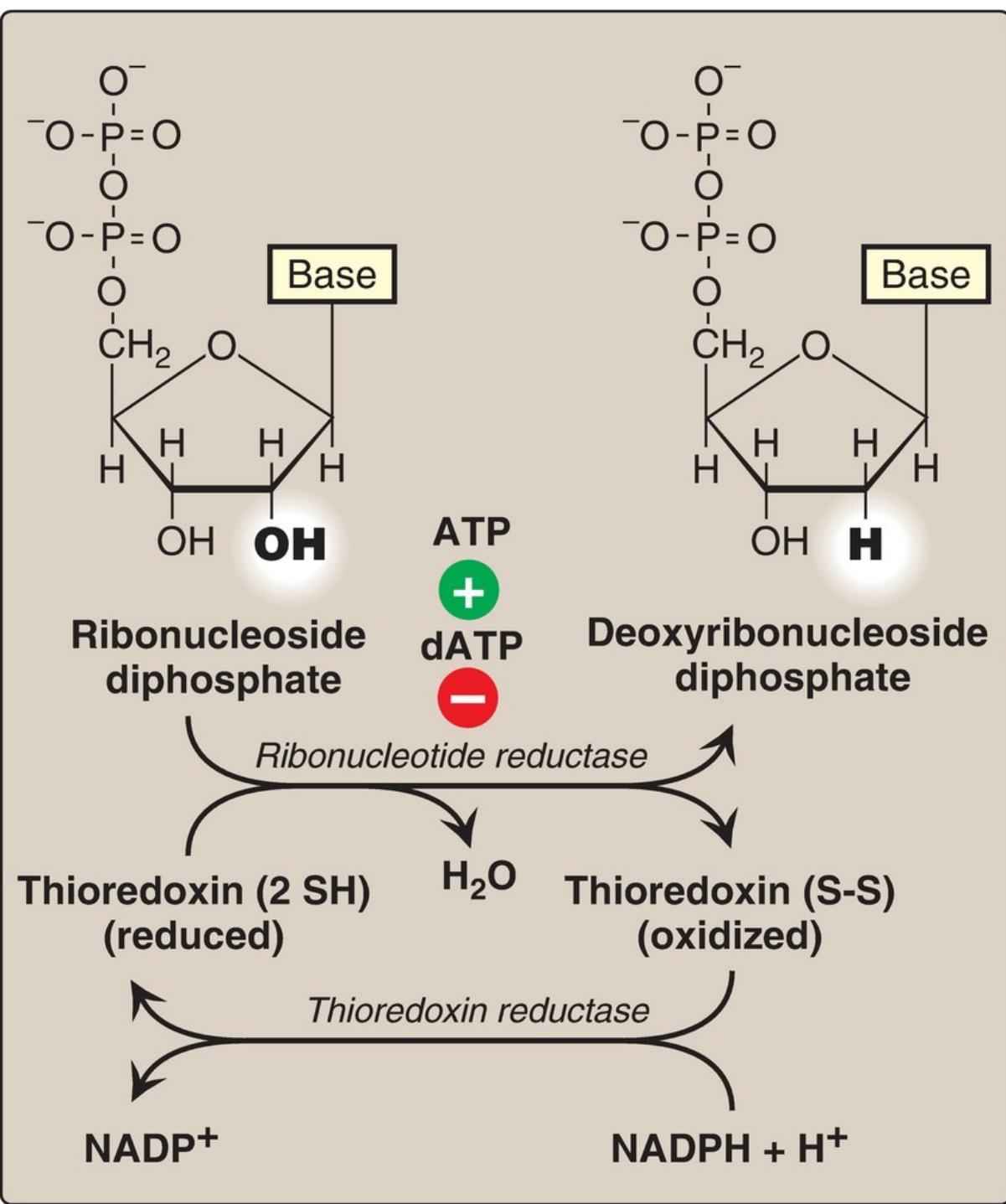


Figure 22.12 Conversion of ribonucleotides to deoxyribonucleotides. NADP(H) = nicotinamide adenine dinucleotide phosphate; dATP = deoxyadenosine triphosphate.

2. Reduced thioredoxin regeneration: Thioredoxin must be converted back

to its reduced form in order to continue performing its function. The reducing equivalents are provided by NADPH + H⁺, and the reaction is catalyzed by ***thioredoxin reductase***, a selenoprotein (see p. 268).

B. Deoxyribonucleotide synthesis regulation

Ribonucleotide reductase is responsible for maintaining a balanced supply of the deoxyribonucleotides required for DNA synthesis. Consequently, the regulation of the enzyme is complex. In addition to the catalytic site, R1 contains two distinct allosteric sites involved in regulating enzymic activity ([Fig. 22.13](#)).

ACTIVITY SITES

ATP activates the enzyme.
dATP inhibits the enzyme.

SUBSTRATE SPECIFICITY SITES

ATP, dATP, dTTP, and dGTP
regulate the reduction of
specific ribonucleotides.

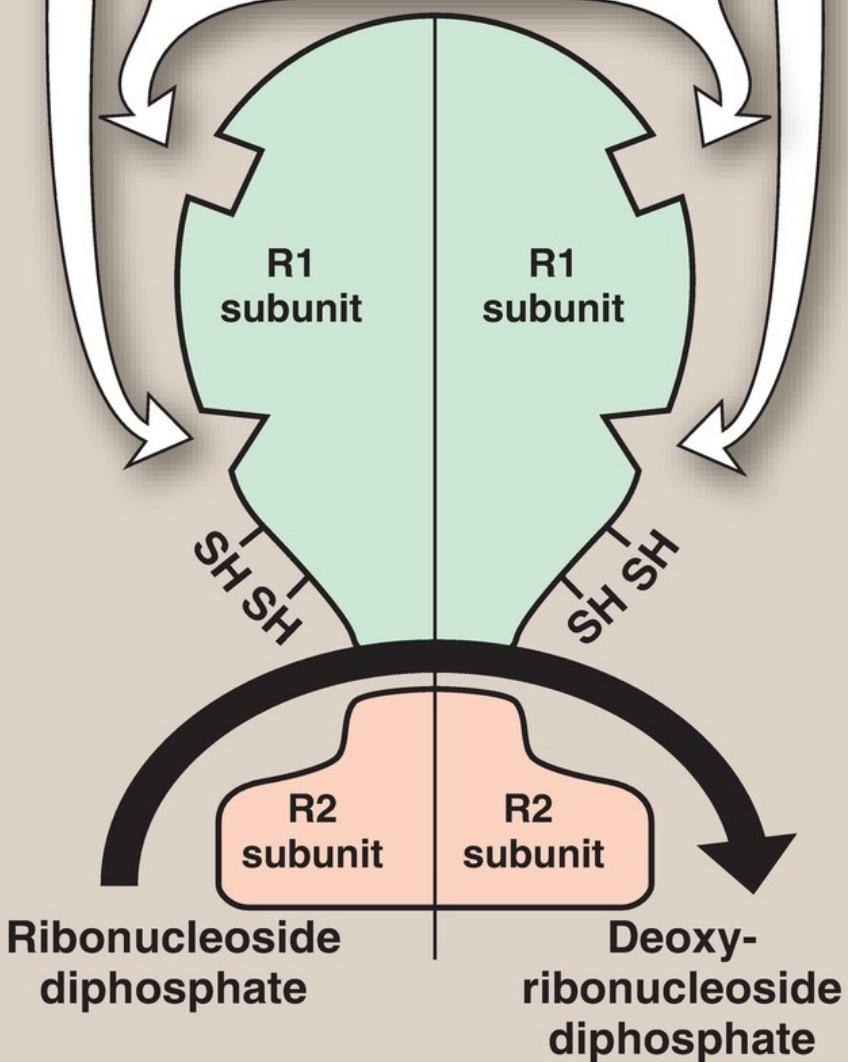


Figure 22.13 Regulation of **ribonucleotide reductase**. dATP, dTTP, and dGTP = deoxyadenosine, deoxythymidine, and deoxyguanosine triphosphates. [Note: The R1 subunit is also referred to as α and the R2 as β .]

1. Activity sites: The binding of dATP to allosteric sites (known as activity sites) on R1 inhibits the overall catalytic activity of the enzyme and, therefore, prevents the reduction of any of the four nucleoside diphosphates. This effectively prevents DNA synthesis and explains the toxicity of increased levels of dATP seen in conditions such as **adenosine deaminase (ADA)** deficiency (see p. 301). In contrast, ATP bound to these sites activates the enzyme.
2. Substrate specificity sites: The binding of nucleoside triphosphates to additional allosteric sites (known as substrate specificity sites) on R1 regulates substrate specificity, causing an increase in the conversion of different species of ribonucleotides to deoxyribonucleotides as they are required for DNA synthesis. For example, deoxythymidine triphosphate binding at the specificity site causes a conformational change that allows reduction of GDP to dGDP at the catalytic site when ATP is at the activity site.

The drug hydroxyurea (hydroxycarbamide) inhibits **ribonucleotide reductase**, thereby inhibiting the generation of substrates for DNA synthesis. The drug is an antineoplastic agent and is used in the treatment of cancers such as melanoma. Hydroxyurea is also used in the treatment of sickle cell anemia (see p. 36). However, the increase in fetal hemoglobin seen with hydroxyurea is because of changes in gene expression and not to **ribonucleotide reductase** inhibition.

V. PURINE NUCLEOTIDE DEGRADATION

Degradation of dietary nucleic acids occurs in the small intestine, where pancreatic **nucleases** hydrolyze them to nucleotides. The nucleotides are sequentially degraded by intestinal enzymes to nucleosides, phosphorylated sugars, and free bases. Uric acid is the end product of intestinal purine degradation. [Note: Purine nucleotides from de novo synthesis are degraded in

the liver primarily. The free bases are sent out from the liver and salvaged by peripheral tissues.]

A. Degradation in the small intestine

Ribonucleases and **deoxyribonucleases**, secreted by the pancreas, hydrolyze dietary RNA and DNA to oligonucleotides that are further hydrolyzed by pancreatic **phosphodiesterases**, producing a mixture of 3'- and 5'-mononucleotides. At the intestinal mucosal surface, **nucleotidases** remove the phosphate groups hydrolytically, releasing nucleosides that are taken into enterocytes by sodium-dependent transporters and degraded by **nucleosidases (nucleoside phosphorylases)** to free bases plus (deoxy) ribose 1-phosphate. Dietary purine bases are not used to any appreciable extent for the synthesis of tissue nucleic acids. Instead, they are degraded to uric acid in the enterocytes. Most of the uric acid enters the blood and is eventually excreted in the urine. A summary of this pathway is shown in [Figure 22.14](#). [Note: Mammals other than primates express **urate oxidase (uricase)**, which cleaves the purine ring, generating allantoin. Modified recombinant **urate oxidase** is now used clinically to lower urate levels.]

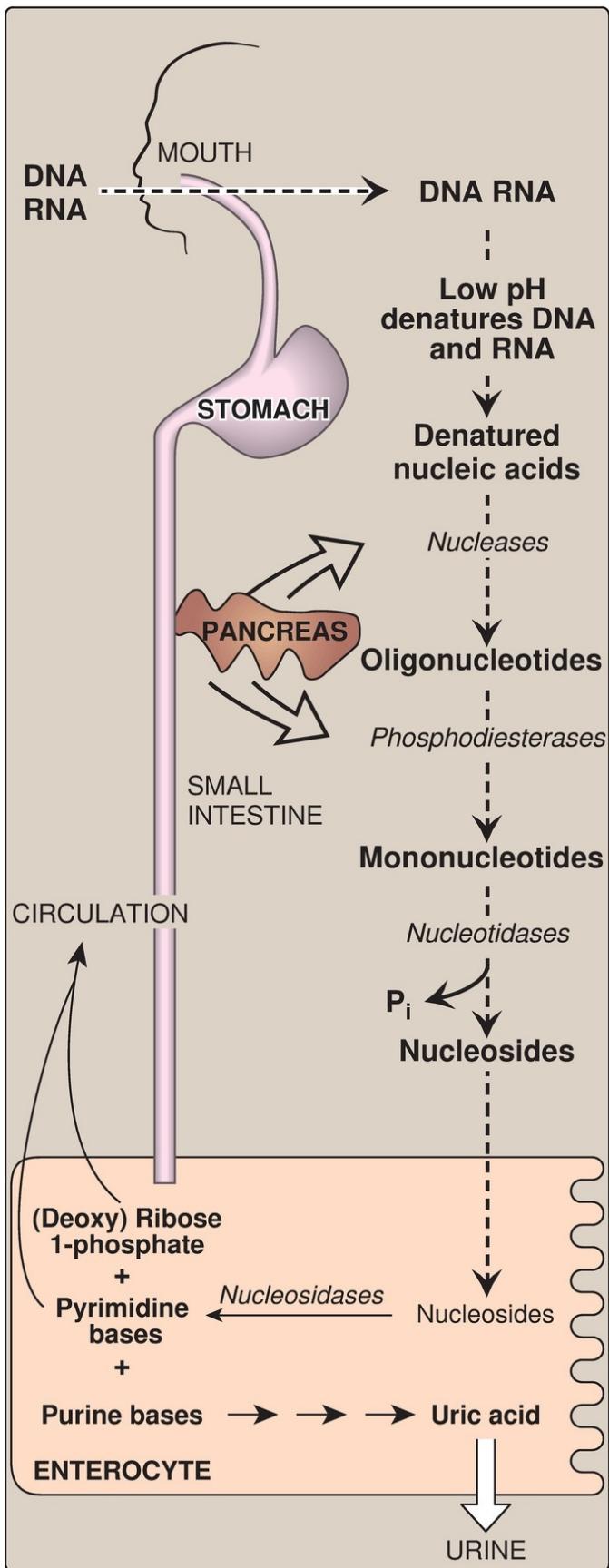


Figure 22.14 Digestion of dietary nucleic acids. P_i = inorganic phosphate.

B. Uric acid formation

A summary of the steps in the production of uric acid and the genetic diseases associated with deficiencies of specific degradative enzymes are shown in [Figure 22.15](#). [Note: The bracketed numbers refer to specific reactions in the figure.]

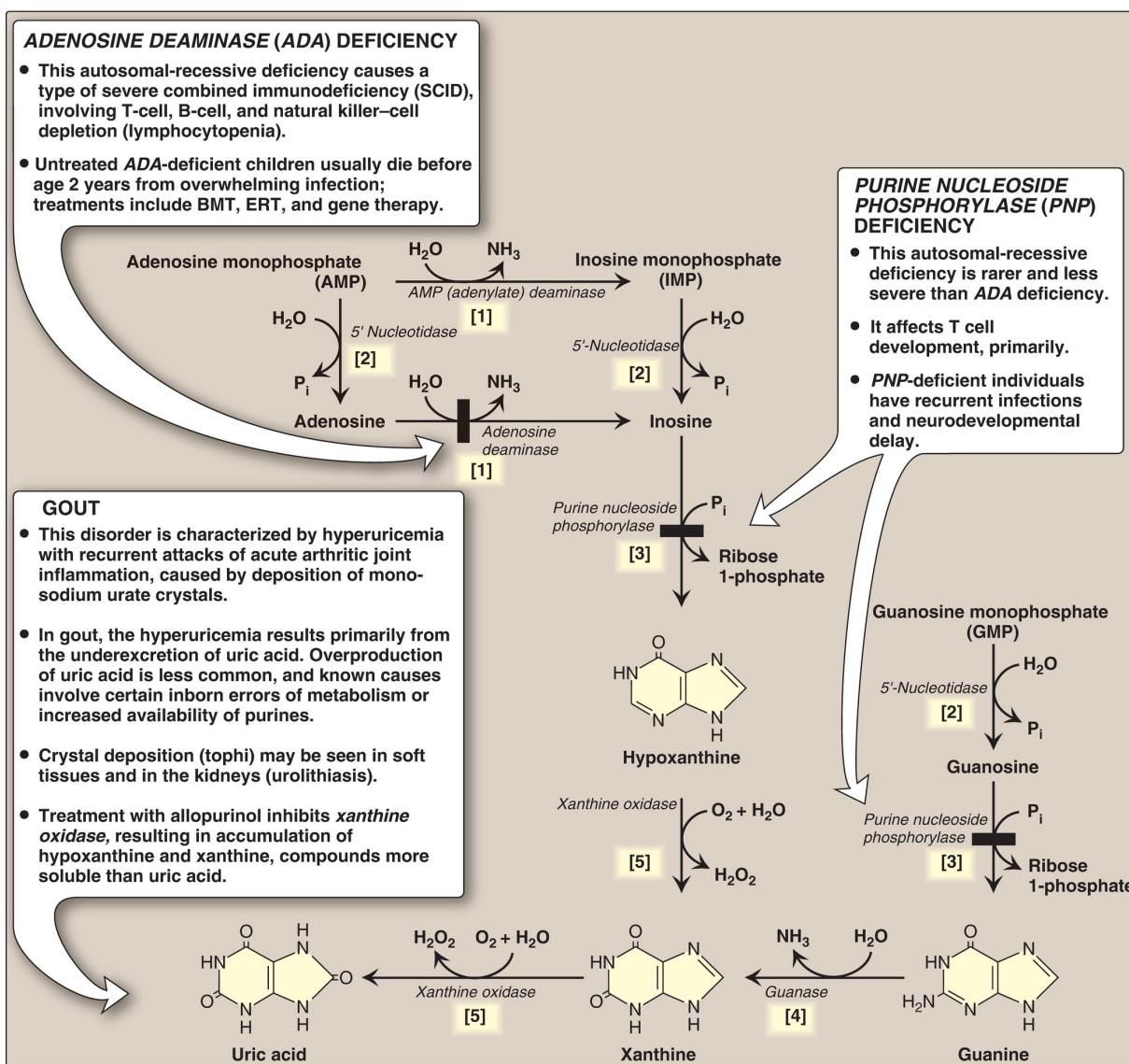


Figure 22.15 The degradation of purine nucleotides to uric acid, illustrating

some of the genetic diseases associated with this pathway. [Note: The numbers in brackets refer to the corresponding numbered citations in the text.] BMT = bone marrow transplantation; ERT = enzyme replacement therapy; P_i = inorganic phosphate; H₂O₂ = hydrogen peroxide; NH₃ = ammonia.

- [1] An amino group is removed from AMP to produce IMP by **AMP (adenylate) deaminase** or from adenosine to produce inosine (hypoxanthine-ribose) by **adenosine deaminase**.
- [2] IMP and GMP are converted into their respective nucleoside forms, inosine and guanosine, by the action of **5'-nucleotidase**.
- [3] **Purine nucleoside phosphorylase** converts inosine and guanosine into their respective purine bases, hypoxanthine and guanine. [Note: A **mutase** interconverts ribose 1- and ribose 5-phosphate.]
- [4] Guanine is deaminated to form xanthine.
- [5] Hypoxanthine is oxidized by molybdenum-containing **xanthine oxidase (XO)** to xanthine, which is further oxidized by **XO** to uric acid, the final product of human purine degradation. Uric acid is excreted primarily in the urine.

C. Diseases associated with purine degradation

1. Gout: Gout is a disorder initiated by high levels of uric acid (the end product of purine catabolism) in blood (hyperuricemia), as a result of either the overproduction or underexcretion of uric acid. The hyperuricemia can lead to the deposition of monosodium urate (MSU) crystals in the joints and an inflammatory response to the crystals, causing first acute and then progressing to chronic gouty arthritis. Nodular masses of MSU crystals (tophi) may be deposited in the soft tissues, resulting in chronic tophaceous gout ([Fig. 22.16](#)). Formation of uric acid stones in the kidney (urolithiasis) may also be seen. [Note: Hyperuricemia is not sufficient to cause gout, but gout is always preceded by hyperuricemia. Hyperuricemia is typically asymptomatic but may be indicative of comorbid conditions such as hypertension.] The definitive diagnosis of gout requires aspiration and examination of synovial fluid ([Fig. 22.17](#)) from an affected joint (or material from a tophus) using polarized light microscopy to confirm the presence of

needle-shaped MSU crystals ([Fig. 22.18](#)).



Figure 22.16 Tophaceous gout.

Arthrocentesis: Joint aspiration is a procedure whereby a sterile needle and syringe are used to drain fluid from a joint.

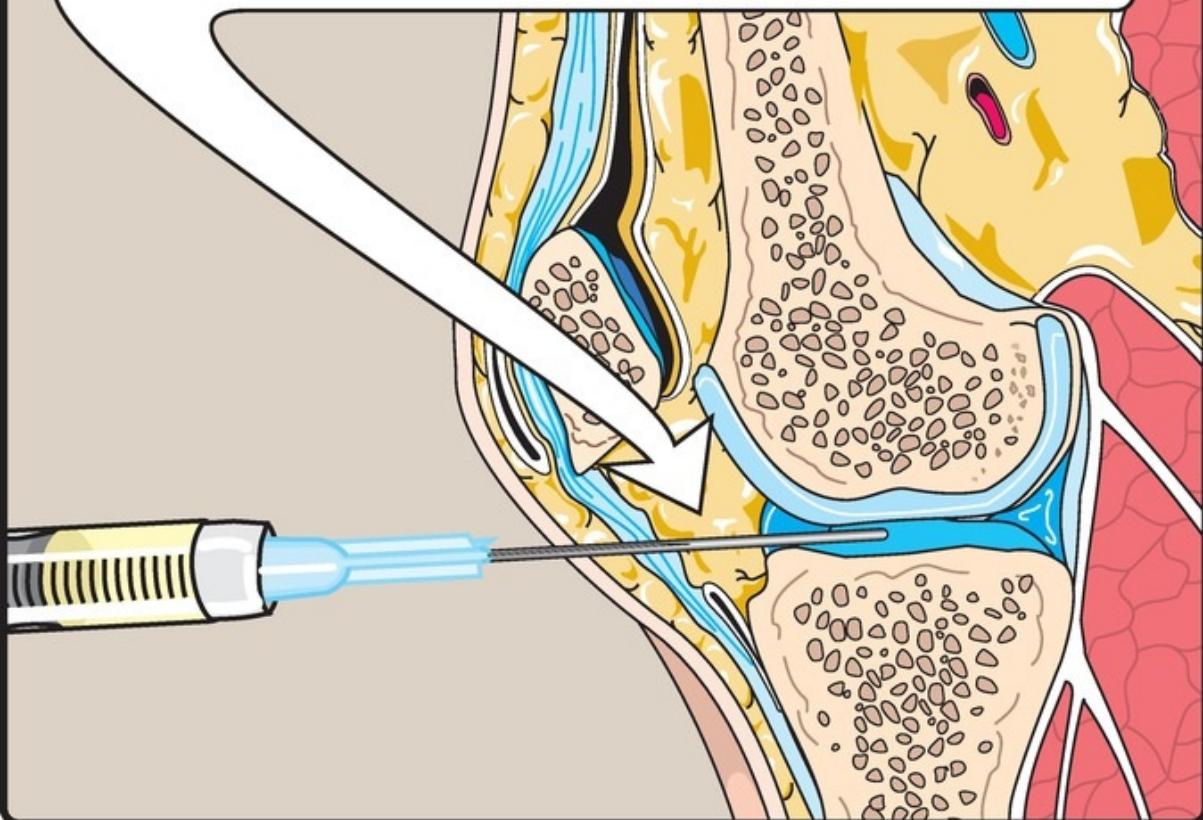


Figure 22.17 Analysis of joint fluid can help to define causes of joint swelling and arthritis, such as infection, gout, and rheumatoid disease.

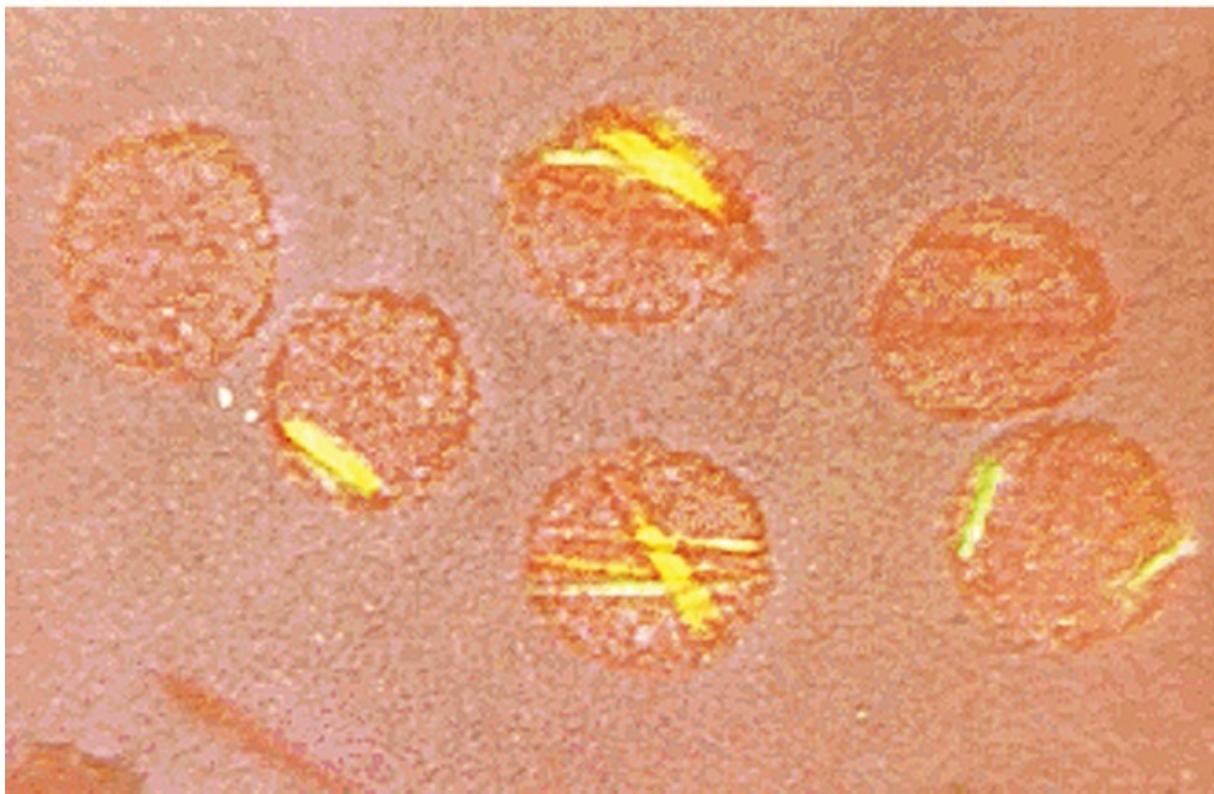


Figure 22.18 Gout can be diagnosed by the presence of negatively birefringent monosodium urate crystals in aspirated synovial fluid examined by polarized light microscopy. Here, crystals are seen within polymorphonuclear leukocytes.

- a. Uric acid underexcretion: In >90% of individuals with hyperuricemia, the cause is underexcretion of uric acid. Underexcretion can be primary, because of as-yet-unidentified inherent excretory defects, or secondary to known disease processes that affect how the kidney handles urate (for example, in lactic acidosis, lactate increases renal urate reabsorption, thereby decreasing its excretion) and to environmental factors such as the use of drugs (for example, thiazide diuretics) or exposure to lead (saturnine gout).
- b. Uric acid overproduction: A less common cause of hyperuricemia is from the overproduction of uric acid. Primary hyperuricemia is, for the most part, idiopathic (having no known cause). However, several identified mutations in the gene for X-linked **PRPP synthetase** result in the enzyme having an increased maximal velocity ($[V_{max}]$ see p. 57) for the production of PRPP, a lower K_m (see p. 59) for ribose 5-phosphate, or a decreased sensitivity to purine nucleotides, its

allosteric inhibitors (see p. 62). In each case, increased availability of PRPP increases purine production, resulting in elevated levels of plasma uric acid. Lesch-Nyhan syndrome (see p. 296) also causes hyperuricemia as a result of the decreased salvage of hypoxanthine and guanine and the subsequent increased availability of PRPP. Secondary hyperuricemia is typically the consequence of increased availability of purines (for example, in patients with myeloproliferative disorders or who are undergoing chemotherapy and so have a high rate of cell turnover). Hyperuricemia can also be the result of seemingly unrelated metabolic diseases, such as von Gierke disease (see Fig. 11.8 on p. 130) or hereditary fructose intolerance (see p. 138).

A diet rich in meat, seafood (particularly shellfish), and ethanol is associated with increased risk of gout, whereas a diet rich in low-fat dairy products is associated with a decreased risk.

- c. Treatment: Acute attacks of gout are treated with anti-inflammatory agents. Colchicine, steroidal drugs such as prednisone, and nonsteroidal drugs such as indomethacin are used. [Note: Colchicine prevents formation of microtubules, thereby decreasing the movement of neutrophils into the affected area. Like the other anti-inflammatory drugs, it has no effect on uric acid levels.] Long-term therapeutic strategies for gout involve lowering the uric acid level below its saturation point (6.5 mg/dl), thereby preventing the deposition of MSU crystals. Uricosuric agents, such as probenecid or sulfinpyrazone, that increase renal excretion of uric acid, are used in patients who are underexcretors of uric acid. Allopurinol, a structural analog of hypoxanthine, inhibits uric acid synthesis and is used in patients who are overproducers of uric acid. Allopurinol is oxidized to oxypurinol, a long-lived inhibitor of **XO**. This results in an accumulation of hypoxanthine and xanthine (see Fig. 22.15), compounds more soluble than uric acid and, therefore, less likely to initiate an inflammatory response. In patients with normal levels of **HGPRT**, the hypoxanthine can be salvaged, reducing the levels of PRPP and, therefore, *de novo* purine synthesis. Febuxostat, a nonpurine inhibitor of **XO**, is also available. [Note: Uric acid levels in the blood normally are close to the saturation point. One reason for this may be the strong antioxidant effects of uric acid.]

2. Adenosine deaminase deficiency

ADA is expressed in a variety of tissues, but, in humans, lymphocytes have the highest activity of this cytoplasmic enzyme. A deficiency of **ADA** results in an accumulation of adenosine, which is converted to its ribonucleotide or deoxyribonucleotide forms by cellular **kinases**. As dATP levels rise, **ribonucleotide reductase** is inhibited, thereby preventing the production of all deoxyribose-containing nucleotides (see p. 297). Consequently, cells cannot make DNA and divide. [Note: The dATP and adenosine that accumulate in **ADA** deficiency lead to developmental arrest and apoptosis of lymphocytes.] In its most severe form, this autosomal-recessive disorder causes a type of severe combined immunodeficiency disease (SCID), involving a decrease in T cells, B cells, and natural killer cells. **ADA** deficiency accounts for ~14% of cases of SCID in the United States. Treatments include bone marrow transplantation, enzyme replacement therapy, and gene therapy (see p. 501). Without appropriate treatment, children with this disorder usually die from infection by age 2 years. [Note: **Purine nucleoside phosphorylase** deficiency results in a less severe immunodeficiency primarily involving T cells.]

VI. PYRIMIDINE SYNTHESIS AND DEGRADATION

Unlike the synthesis of the purine ring, which is constructed on a pre-existing ribose 5-phosphate, the pyrimidine ring is synthesized before being attached to ribose 5-phosphate, which is donated by PRPP. The sources of the atoms in the pyrimidine ring are glutamine, CO₂, and aspartate (Fig. 22.19).

Amide nitrogen (R-group) of glutamine

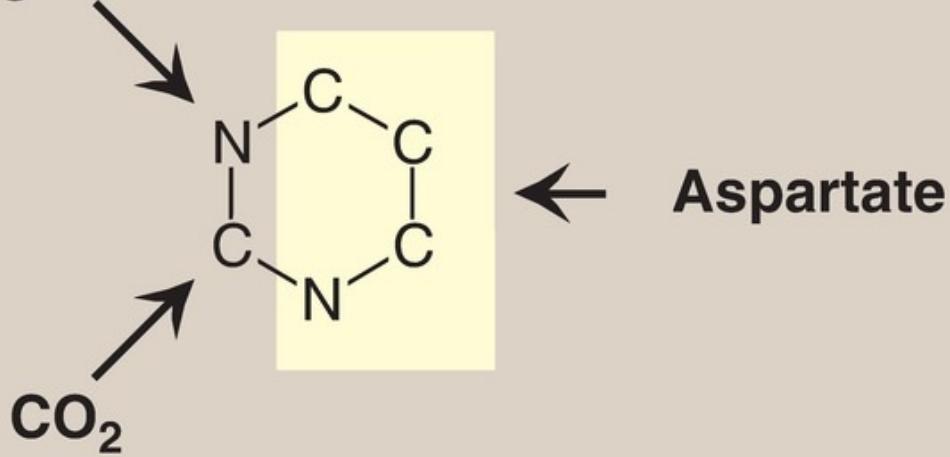


Figure 22.19 Sources of the individual atoms in the pyrimidine ring. CO₂ = carbon dioxide.

A. Carbamoyl phosphate synthesis

The regulated step of this pathway in mammalian cells is the synthesis of carbamoyl phosphate from glutamine and CO₂, catalyzed by **carbamoyl phosphate synthetase (CPS) II**. CPS II is inhibited by uridine triphosphate (the end product of this pathway, which can be converted into the other pyrimidine nucleotides) and is activated by PRPP. [Note: Carbamoyl phosphate, synthesized by **CPS I**, is also a precursor of urea (see p. 253). Defects in **ornithine transcarbamylase** of the urea cycle promote pyrimidine synthesis because of increased availability of carbamoyl phosphate. A comparison of the two enzymes is presented in [Figure 22.20](#).]

| Variable | <i>CPS I</i> | <i>CPS II</i> |
|--------------------|----------------------------------|---|
| Cellular location | Mitochondria | Cytosol |
| Pathway involved | Urea cycle | Pyrimidine synthesis |
| Source of nitrogen | Ammonia | γ -Amide group of glutamine |
| Regulators | Activator: N-acetyl-glutamate | Activator: PRPP Inhibitor: UTP |

Figure 22.20 Summary of the differences between *carbamoyl phosphate synthetase (CPS) I* and *II*. PRPP = 5-phosphoribosyl-1-pyrophosphate; UTP = uridine triphosphate.

B. Orotic acid synthesis

The second step in pyrimidine synthesis is the formation of carbamoylaspartate, catalyzed by *aspartate transcarbamoylase*. The pyrimidine ring is then closed by *dihydroorotase*. The resulting dihydroorotate is oxidized to produce orotic acid (orotate), as shown in Figure 22.21. The human enzyme that produces orotate, *dihydroorotate dehydrogenase*, is a flavin mononucleotide-containing protein of the inner

mitochondrial membrane. All other enzymes in pyrimidine biosynthesis are cytosolic. [Note: The first three enzymic activities in this pathway (**CPS II**, **aspartate transcarbamoylase**, and **dihydroorotase**) are actually three different catalytic domains of a single polypeptide known as **CAD** from the first letter in the name of each domain. (See p. 18 for a discussion of domains.) This is an example of a multifunctional or multicatalytic polypeptide that facilitates the ordered synthesis of an important compound. Synthesis of the purine nucleotide IMP also involves multifunctional proteins.]

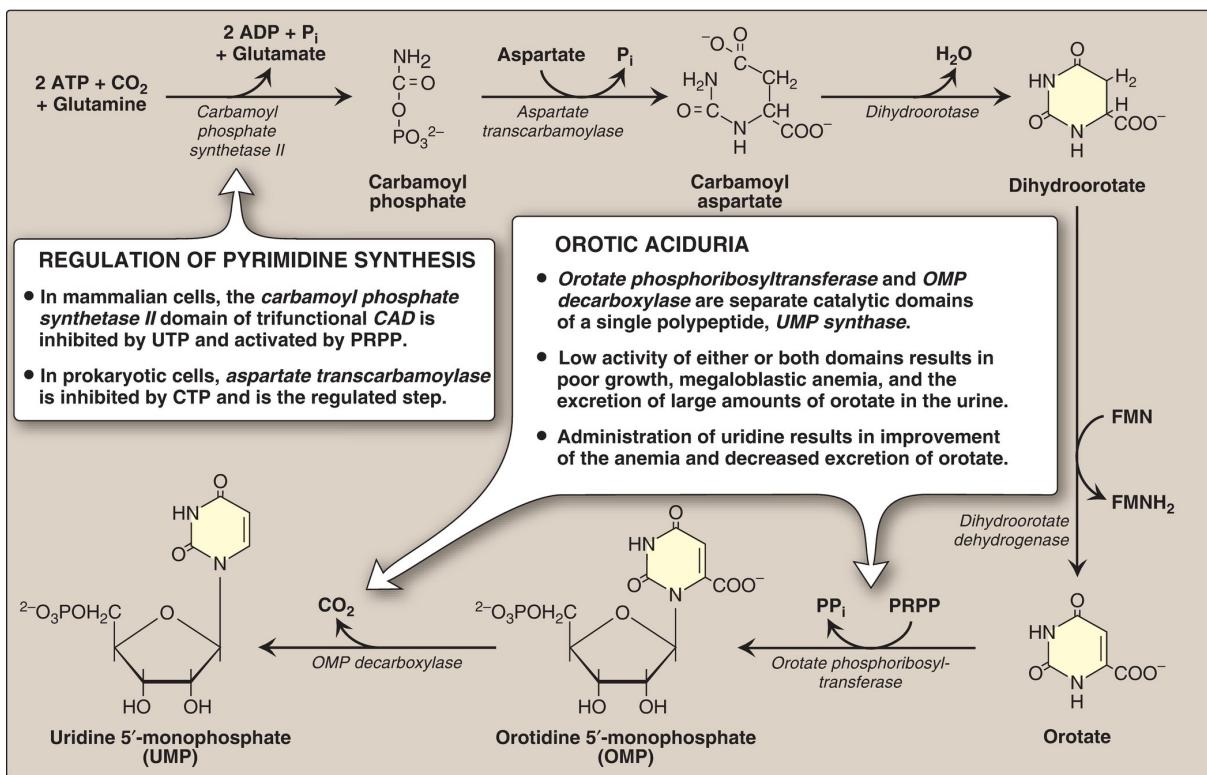


Figure 22.21 *De novo* pyrimidine synthesis. ADP = adenosine diphosphate; P_i = inorganic phosphate; FMN(H₂) = flavin mononucleotide; CTP = cytidine triphosphate; PRPP = 5-phosphoribosyl-1-pyrophosphate; PP_i = pyrophosphate.

C. Pyrimidine nucleotide synthesis

The completed pyrimidine ring is converted to the nucleotide orotidine monophosphate (OMP) in the second stage of pyrimidine nucleotide synthesis (see Fig. 22.21). As seen with the purines, PRPP is the ribose 5-

phosphate donor. The enzyme ***orotate phosphoribosyltransferase*** produces OMP and releases pyrophosphate, thereby making the reaction biologically irreversible. [Note: Both purine and pyrimidine synthesis require glutamine, aspartic acid, and PRPP as essential precursors.] OMP (orotidylate) is decarboxylated to uridine monophosphate (UMP) by ***orotidylate decarboxylase***. The ***phosphoribosyltransferase*** and ***decarboxylase*** activities are separate catalytic domains of a single polypeptide called ***UMP synthase***. Hereditary orotic aciduria (a very rare disorder) may be caused by a deficiency of one or both activities of this bifunctional enzyme, resulting in orotic acid in the urine (see [Fig. 22.21](#)). UMP is sequentially phosphorylated to UDP and UTP. [Note: The UDP is a substrate for ***ribonucleotide reductase***, which generates dUDP. The dUDP is phosphorylated to dUTP, which is rapidly hydrolyzed to dUMP by ***UTP diphosphatase (dUTPase)***. Thus, ***dUTPase*** plays an important role in reducing availability of dUTP as a substrate for DNA synthesis, thereby preventing erroneous incorporation of uracil into DNA.]

D. Cytidine triphosphate synthesis

Cytidine triphosphate (CTP) is produced by amination of UTP by ***CTP synthetase*** ([Fig. 22.22](#)), with glutamine providing the nitrogen. Some of this CTP is dephosphorylated to CDP, which is a substrate for ***ribonucleotide reductase***. The dCDP product can be phosphorylated to dCTP for DNA synthesis or dephosphorylated to dCMP that is deaminated to dUMP.

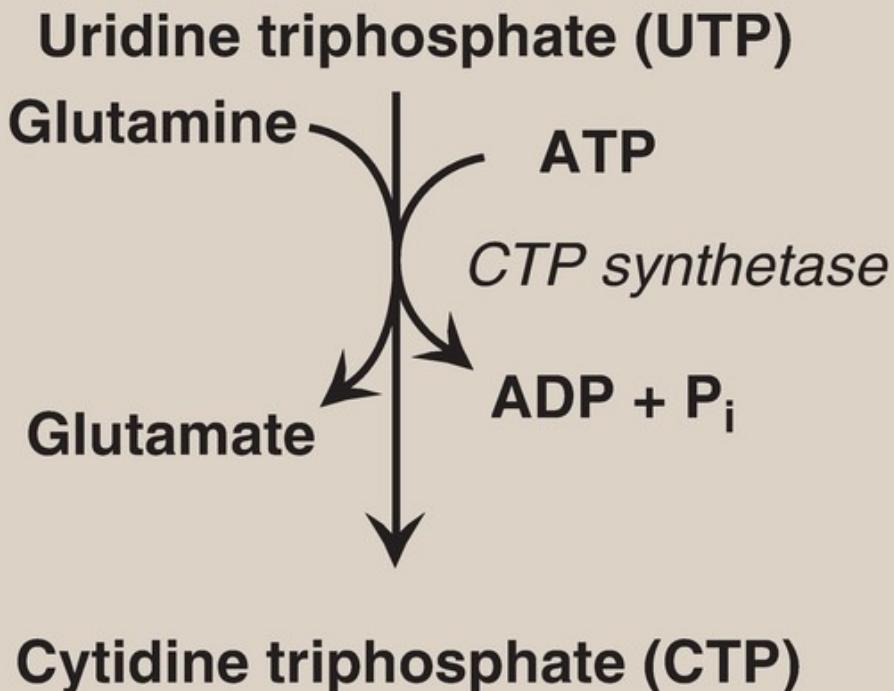


Figure 22.22 Synthesis of CTP from UTP. [Note: CTP, required for RNA synthesis, is converted to dCTP for DNA synthesis.] ADP = adenosine diphosphate; P_i = inorganic phosphate.

E. Deoxythymidine monophosphate synthesis

dUMP is converted to deoxythymidine monophosphate (dTMP) by **thymidylate synthase**, which uses N⁵,N¹⁰-methylene-THF as the source of the methyl group (see p. 267). This is an unusual reaction in that THF contributes not only a one-carbon unit but also two hydrogen atoms from the pteridine ring, resulting in the oxidation of THF to dihydrofolate ([DHF], Fig. 22.23). Inhibitors of **thymidylate synthase** include thymine analogs such as 5-fluorouracil, which serve as antitumor agents. 5-Fluorouracil is metabolically converted to 5-fluorodeoxyuridine monophosphate (5-FdUMP), which becomes permanently bound to the inactivated **thymidylate synthase**, making the drug a suicide inhibitor (see p. 60). DHF can be reduced to THF by **dihydrofolate reductase** (see Fig.

28.2, p. 378), an enzyme that is inhibited by folate analogs such as methotrexate. By decreasing the supply of THF, these drugs not only inhibit purine synthesis (see Fig. 22.7), but, by preventing methylation of dUMP to dTMP, they also decrease the availability of this essential component of DNA. DNA synthesis is inhibited and cell growth slowed. Thus, these drugs are used to treat cancer. [Note: Acyclovir (a purine analog) and AZT (3'-azido-3'-deoxythymidine, a pyrimidine analog) are used to treat infections of herpes simplex virus and human immunodeficiency virus, respectively. Each inhibits the viral **DNA polymerase**.]

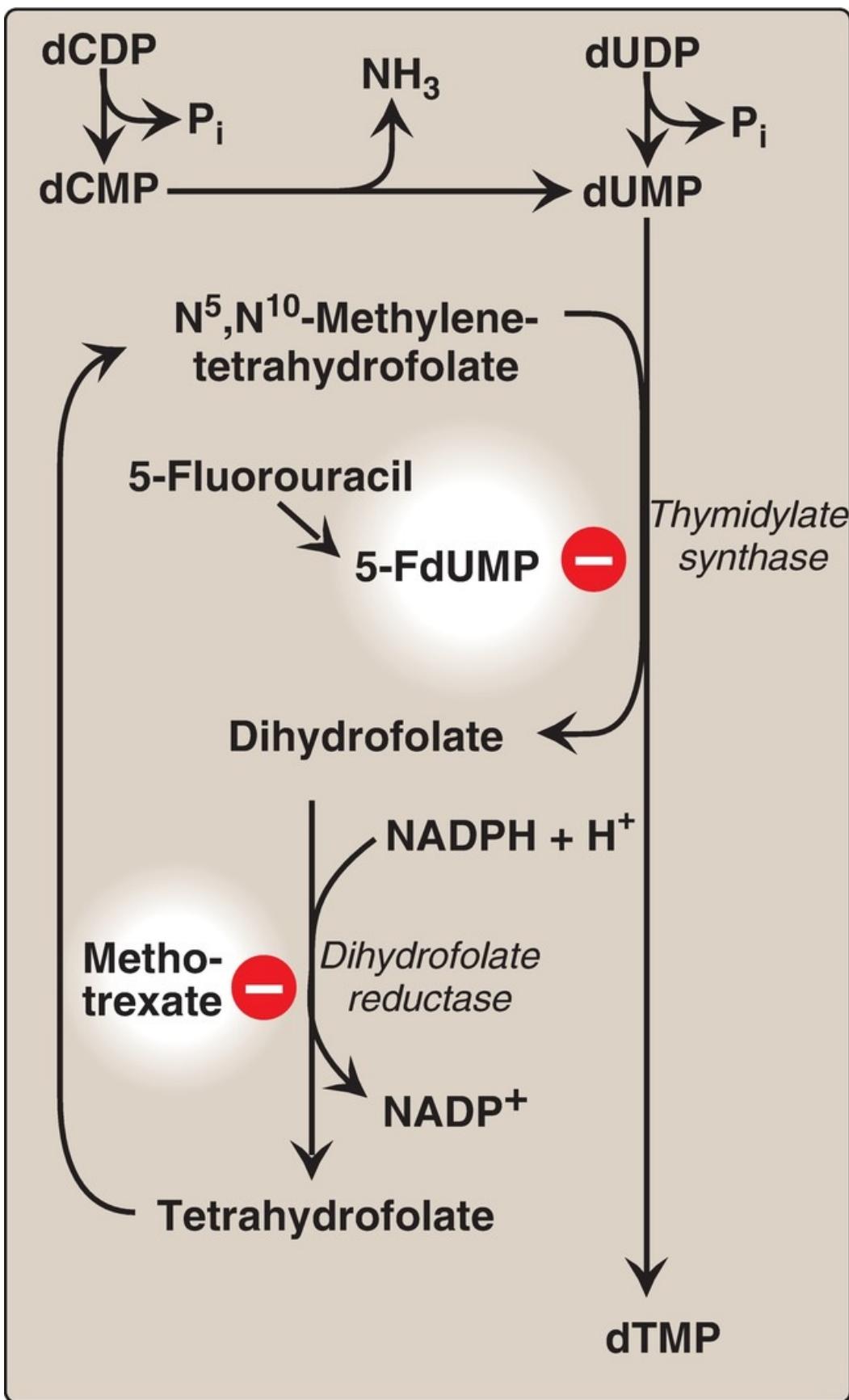


Figure 22.23 Synthesis of dTMP from dUMP, illustrating sites of action of antineoplastic drugs.

F. Pyrimidine salvage and degradation

Unlike the purine ring, which is not cleaved in humans and is excreted as poorly soluble uric acid, the pyrimidine ring is opened and degraded to highly soluble products, β -alanine (from the degradation of CMP and UMP) and β -aminoisobutyrate (from TMP degradation), with the production of ammonia and CO_2 . Pyrimidine bases can be salvaged to nucleosides, which are phosphorylated to nucleotides. However, their high solubility makes pyrimidine salvage less significant clinically than purine salvage. [Note: The salvage of pyrimidine nucleosides is the basis for using uridine in the treatment of hereditary orotic aciduria (see p. 302).]

VII. CHAPTER SUMMARY

Nucleotides are composed of a nitrogenous base (adenine = A, guanine = G, cytosine = C, uracil = U, and thymine = T); a pentose sugar; and one, two, or three phosphate groups (Fig. 22.24). A and G are purines, and C, U, and T are pyrimidines. If the sugar is ribose, the nucleotide is a ribonucleoside phosphate (for example, adenosine monophosphate [AMP]), and it can have several functions in the cell, including being a component of RNA. If the sugar is deoxyribose, the nucleotide is a deoxyribonucleoside phosphate (for example, deoxyAMP) and will be found almost exclusively as a component of DNA. The committed step in purine synthesis uses 5-phosphoribosyl-1-pyrophosphate ([PRPP], an activated pentose that provides the ribose 5-phosphate for *de novo* purine and pyrimidine synthesis and salvage) and nitrogen from glutamine to produce phosphoribosylamine. The enzyme is **glutamine:phosphoribosylpyrophosphate amidotransferase** and is inhibited by AMP and guanosine monophosphate (the end products of the pathway) and activated by PRPP. Purine nucleotides can also be produced from preformed purine bases by using salvage reactions catalyzed by **adenine phosphoribosyltransferase (APRT)** and **hypoxanthine–guanine phosphoribosyltransferase (HGPRT)**. A near-total deficiency of **HGPRT** causes Lesch-Nyhan syndrome, a severe, inherited form of hyperuricemia accompanied by compulsive self-mutilation. All deoxyribonucleotides are synthesized from ribonucleotides by the enzyme **ribonucleotide reductase**. This enzyme is highly regulated (for example, it is strongly inhibited by deoxyadenosine triphosphate [dATP], a compound that is overproduced in bone marrow cells in individuals with **adenosine deaminase (ADA)** deficiency). **ADA** deficiency causes severe combined immunodeficiency disease. The end product of purine degradation is uric acid, a compound of low solubility whose overproduction or undersecretion causes hyperuricemia that, if accompanied by the deposition of monosodium urate crystals in joints and soft tissues and an inflammatory response to those crystals, results in gout. The first step in pyrimidine synthesis, the production of carbamoyl phosphate by **carbamoyl phosphate synthetase II**, is the regulated step in this pathway (it is inhibited by uridine triphosphate

[UTP] and activated by PRPP). The UTP produced by this pathway can be converted to cytidine triphosphate. Deoxyuridine monophosphate can be converted to deoxythymidine monophosphate by ***thymidylate synthase***, an enzyme targeted by anticancer drugs such as 5-fluorouracil. The regeneration of tetrahydrofolate from dihydrofolate produced in the ***thymidylate synthase*** reaction requires ***dihydrofolate reductase***, an enzyme targeted by the drug methotrexate. Pyrimidine degradation results in soluble products.

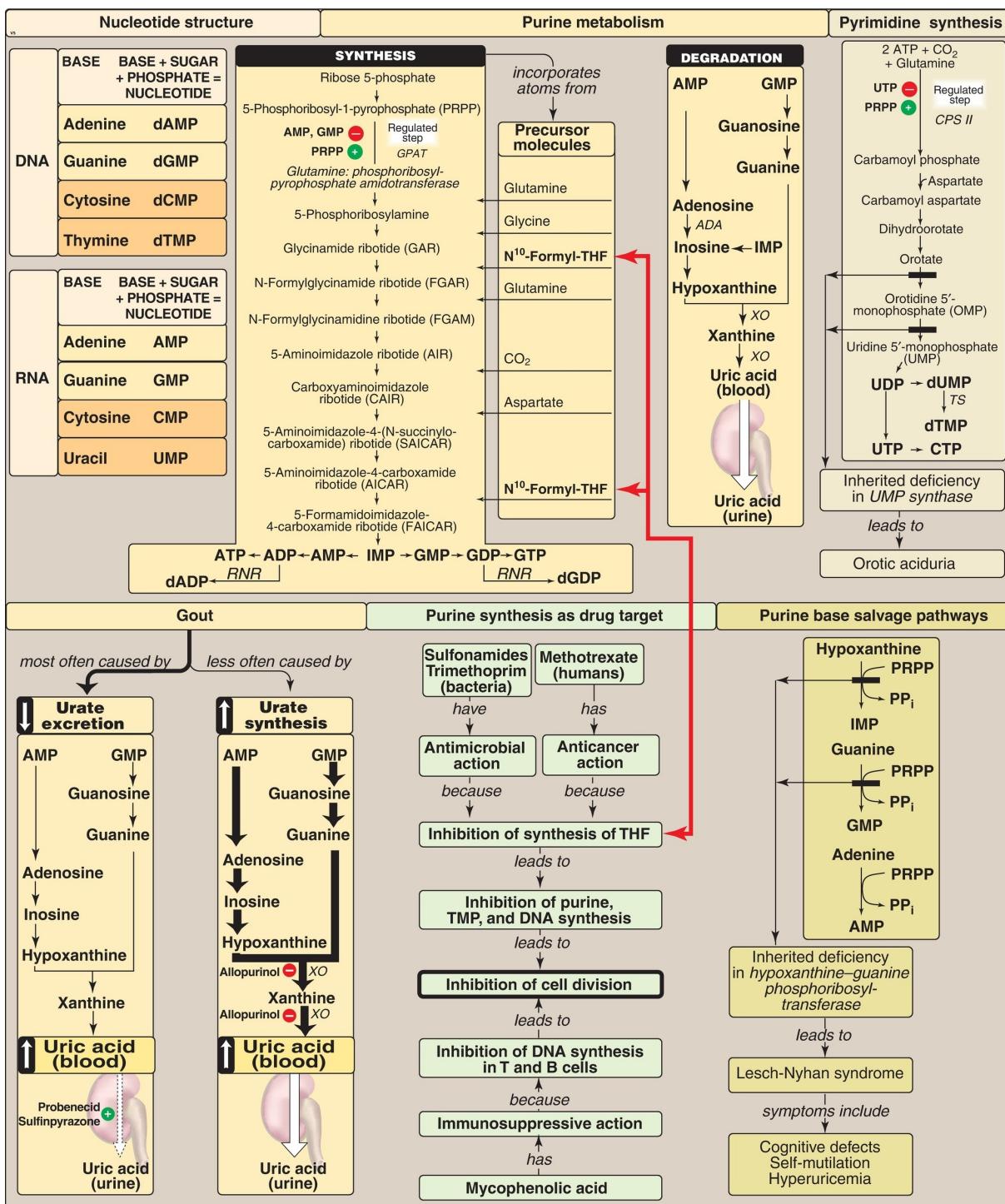


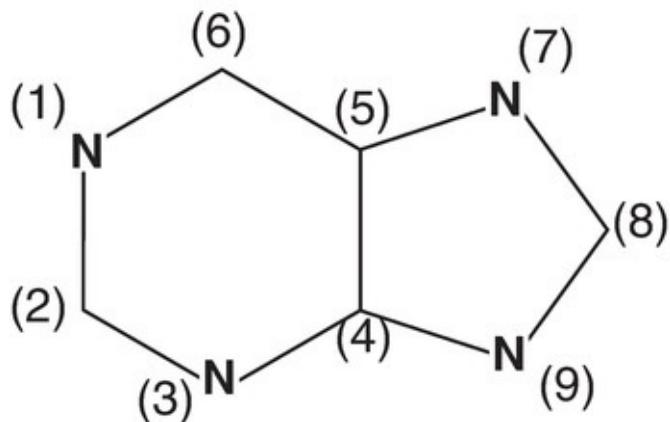
Figure 22.24 Key concept map for nucleotide metabolism. THF = tetrahydrofolate; **GPAT** = *glutamine:phosphoribosylpyrophosphate amidotransferase*; **ADA** = *adenosine deaminase*; **XO** = *xanthine oxidase*; **TS** = *thymidylate synthase*; **RNR** = *ribonucleotide reductase*; **CPS II** = *carbamoyl phosphate synthetase II*; AMP, GMP, CMP, TMP, and IMP = adenosine,

guanosine, cytidine, thymidine, and inosine monophosphates; d = deoxy; PP_i = pyrophosphate; PRPP = 5-phosphoribosyl-1-pyrophosphate.

Study Questions

Choose the ONE best answer.

- 2.1. Azaserine, a drug with research applications, inhibits glutamine-dependent enzymes. Incorporation of which of the ring nitrogens (N) in the generic purine structure shown would most likely be affected by azaserine?



- A. 1
- B. 3
- C. 7
- D. 9

Correct answer = D. The N at position 9 is supplied by glutamine in the first step of purine *de novo* synthesis, and its incorporation would be affected by azaserine. The N at position 1 is supplied by aspartate and at position 7 by glycine. The N at position 3 is also supplied by glutamine, but azaserine would have inhibited purine synthesis prior to this step.

- 2.2. A 42-year-old male patient undergoing radiation therapy for prostate cancer develops severe pain in the metatarsal phalangeal joint of his right big toe. Monosodium urate crystals are detected by polarized light microscopy in

fluid obtained from this joint by arthrocentesis. This patient's pain is directly caused by the overproduction of the end product of which of the following metabolic pathways?

- A. De novo pyrimidine biosynthesis
- B. Pyrimidine degradation
- C. De novo purine biosynthesis
- D. Purine salvage
- E. Purine degradation

Correct answer = E. The patient's pain is caused by gout, resulting from an inflammatory response to the crystallization of excess urate (as monosodium urate) in his joints. Radiation therapy caused cell death, with degradation of nucleic acids and their constituent purines. Uric acid, the end product of purine degradation, is a relatively insoluble compound that can cause gout (and kidney stones). Pyrimidine metabolism is not associated with uric acid production. Overproduction of purines can indirectly result in hyperuricemia. Purine salvage decreases uric acid production.

2.3. Which one of the following enzymes of nucleotide metabolism is correctly paired with its pharmacologic inhibitor?

- A. Dihydrofolate reductase—methotrexate
- B. Inosine monophosphate dehydrogenase—hydroxyurea
- C. Ribonucleotide reductase—5-fluorouracil
- D. Thymidylate synthase—allopurinol
- E. Xanthine oxidase—probenecid

Correct answer = A. Methotrexate interferes with folate metabolism by acting as a competitive inhibitor of the enzyme dihydrofolate reductase. This starves cells for tetrahydrofolate and makes them unable to synthesize purines and thymidine monophosphate. Inosine monophosphate dehydrogenase is inhibited by mycophenolic acid. Ribonucleotide reductase is inhibited by hydroxyurea. Thymidylate synthase is inhibited by 5-fluorouracil. Xanthine oxidase is inhibited by allopurinol. Probenecid increases renal excretion of urate but does not inhibit its production.

2.4. A 1-year-old female patient is lethargic, weak, and anemic. Her height and

weight are low for her age. Her urine contains an elevated level of orotic acid. Activity of uridine monophosphate synthase is low. Administration of which of the following is most likely to alleviate her symptoms?

- A. Adenine
- B. Guanine
- C. Hypoxanthine
- D. Thymidine
- E. Uridine

Correct answer = E. The elevated excretion of orotic acid and low activity of uridine monophosphate (UMP) synthase indicate that the patient has orotic aciduria, a very rare genetic disorder affecting *de novo* pyrimidine synthesis. Deficiencies in one or both catalytic domains of UMP synthase leave the patient unable to synthesize pyrimidines. Uridine, a pyrimidine nucleoside, is a useful treatment because it bypasses the missing activities and can be salvaged to UMP, which can be converted to all the other pyrimidines. Although thymidine is a pyrimidine nucleoside, it cannot be converted to other pyrimidines. Hypoxanthine, guanine, and adenine are all purine bases and cannot be converted to pyrimidines.

2.5. What laboratory test would help in distinguishing an orotic aciduria caused by ornithine transcarbamylase deficiency from that caused by uridine monophosphate synthase deficiency?

Blood ammonia level would be expected to be elevated in ornithine transcarbamylase deficiency that affects the urea cycle but not in uridine monophosphate synthase deficiency.